

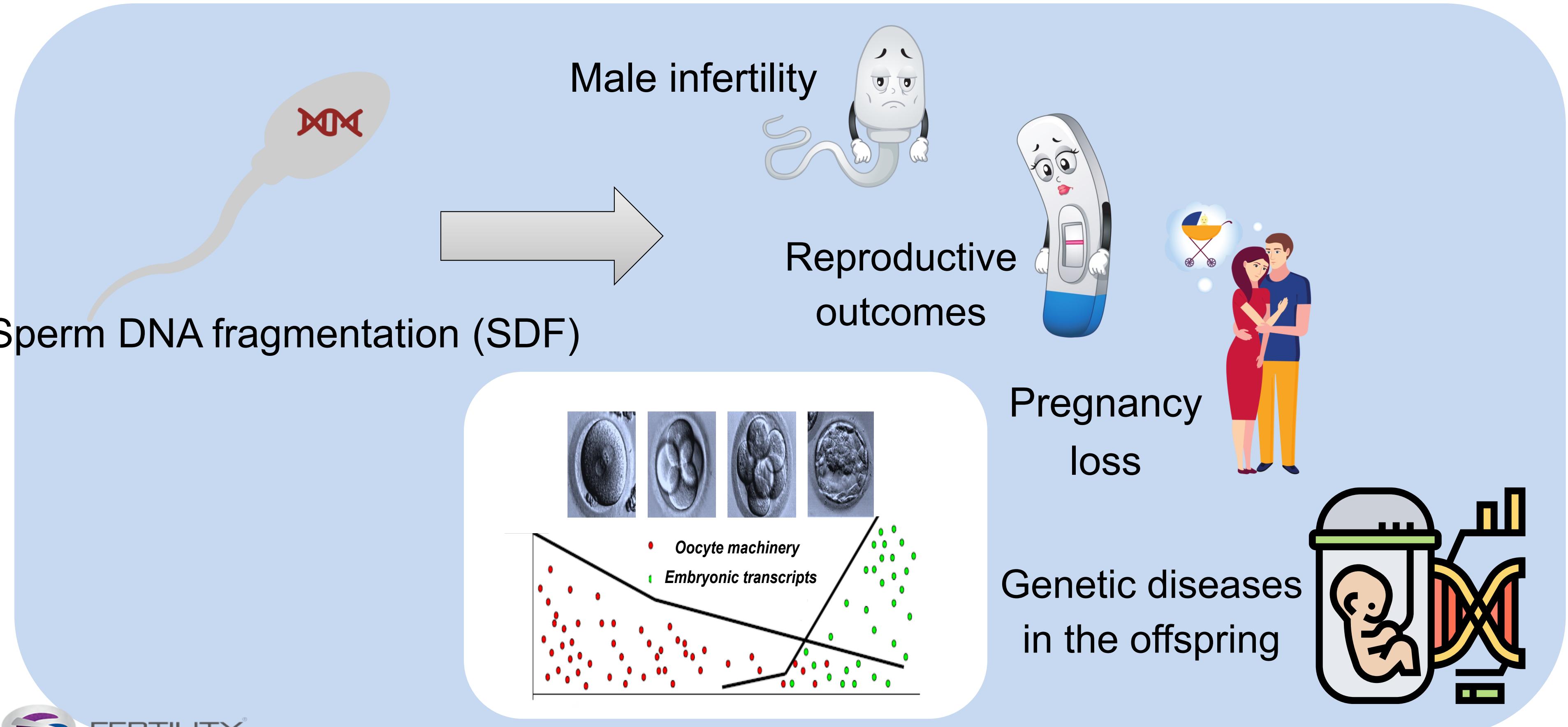


FERTILITY
MEDICAL GROUP

HIGH SPERM DNA FRAGMENTATION INDEX NEGATIVELY IMPACTS EMBRYO MORPHOKINETICS, BUT NOT EMBRYO MORPHOLOGY AND DEVELOPMENT RATES: THE IMPORTANCE OF TIME-LAPSE IMAGING SYSTEM

Edson Borges Jr., Amanda Setti, Daniela Paes de Almeida Ferreira Braga,
Rodrigo Rosa Provenza, Patrícia Guilherme, Assumpto Iaconelli Jr.

INTRODUCTION



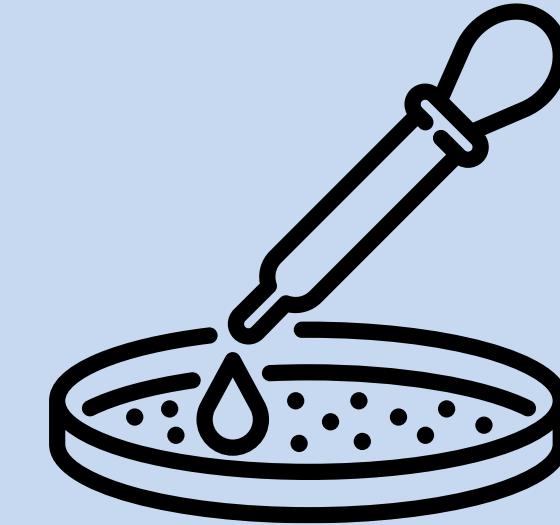
INTRODUCTION



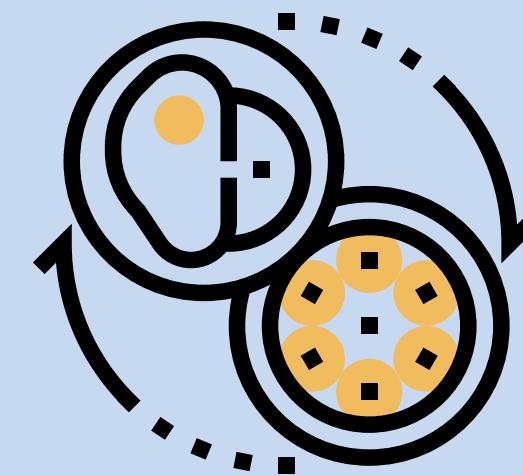
Parental age



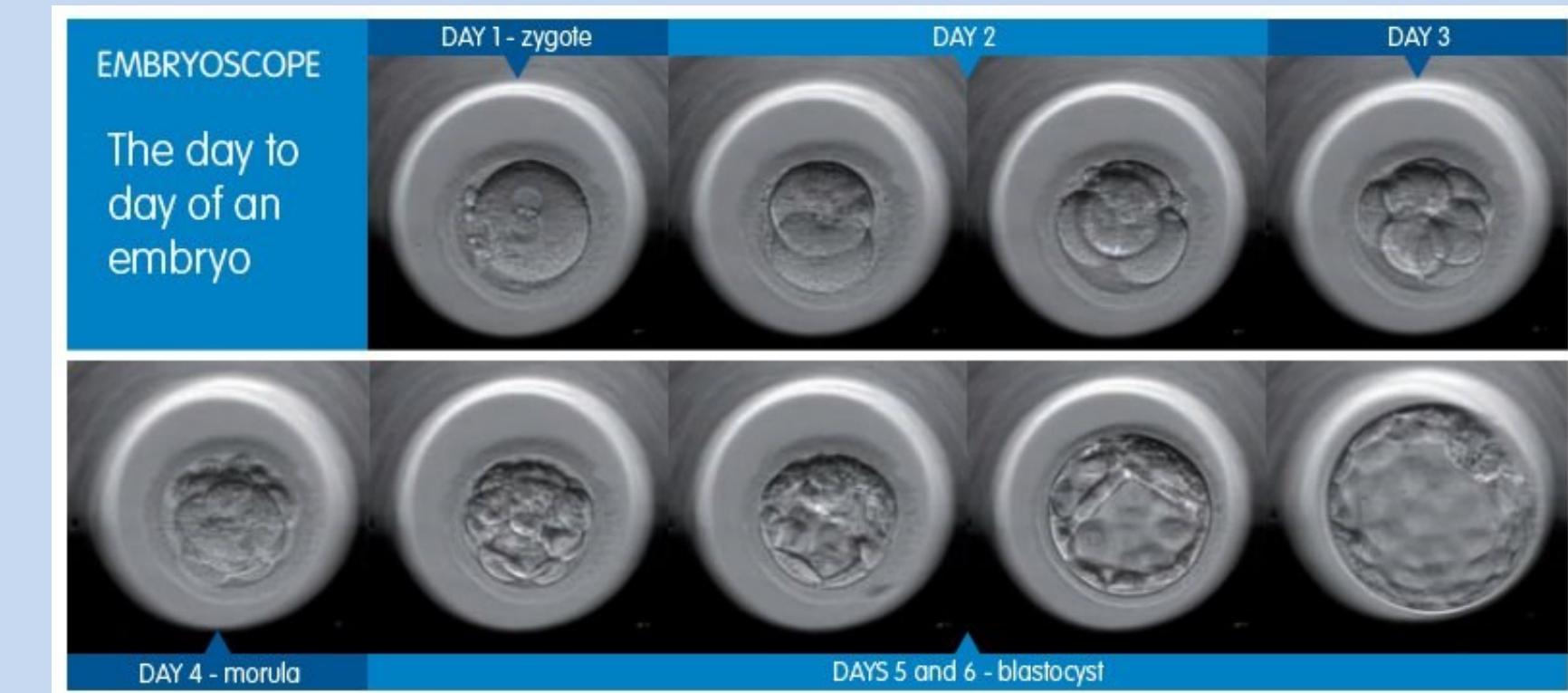
COS



Culture media



Intrinsic embryonic quality



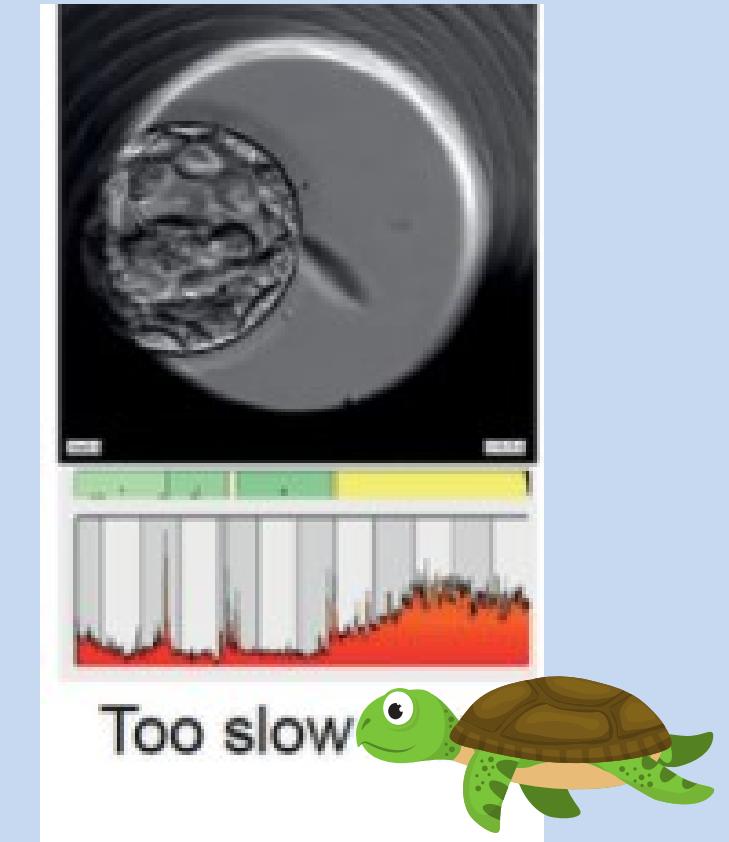
Time-lapse imaging (TLI)

INTRODUCTION



Hypothesis

DNA fragmentation may interfere with speed and pattern of cell divisions

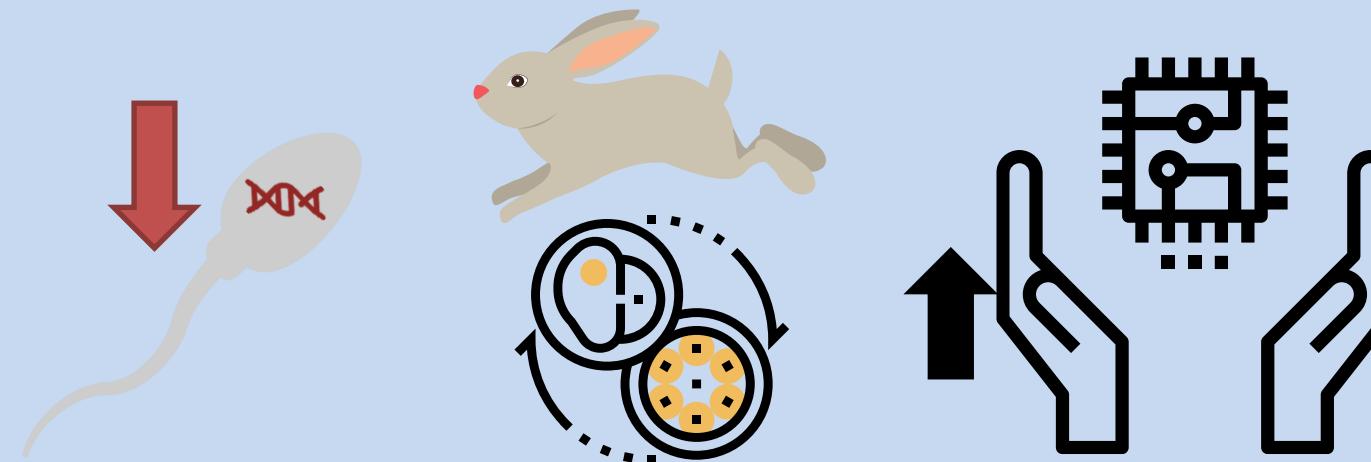


INTRODUCTION

➤ Reprod Biol. 2015 Jun;15(2):94-100. doi: 10.1016/j.repbio.2015.03.003. Epub 2015 Apr 2.

The effect of sperm DNA fragmentation on the dynamics of the embryonic development in intracytoplasmatic sperm injection

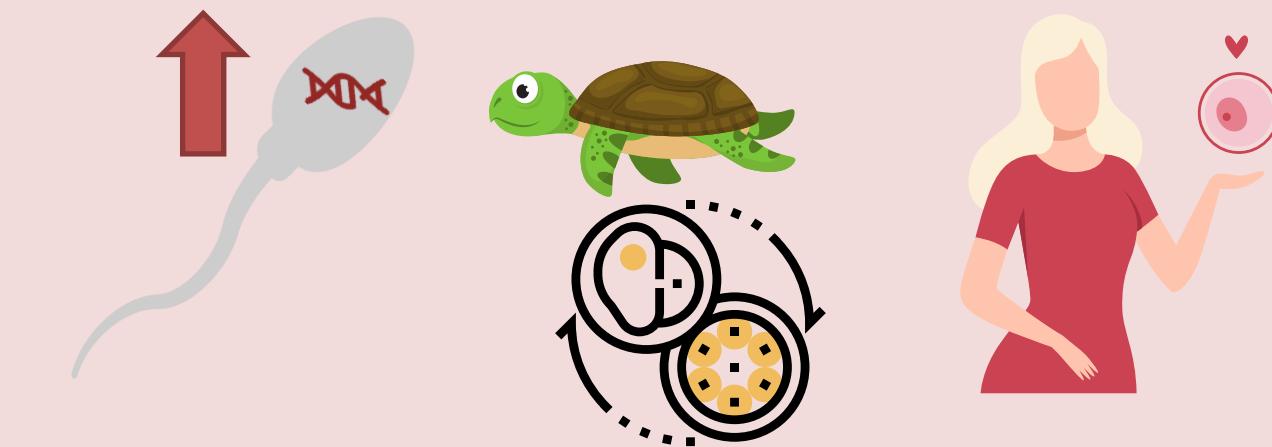
Artur Wdowiak ¹, Szymon Bakalczuk ², Grzegorz Bakalczuk ³



➤ Andrology. 2018 Sep;6(5):697-706. doi: 10.1111/andr.12551. Epub 2018 Sep 26.

High sperm DNA fragmentation delays human embryo kinetics when oocytes from young and healthy donors are microinjected

M Esbert ¹, A Pacheco ², S R Soares ³, D Amorós ¹, M Florensa ¹, A Ballesteros ¹, M Meseguer ⁴

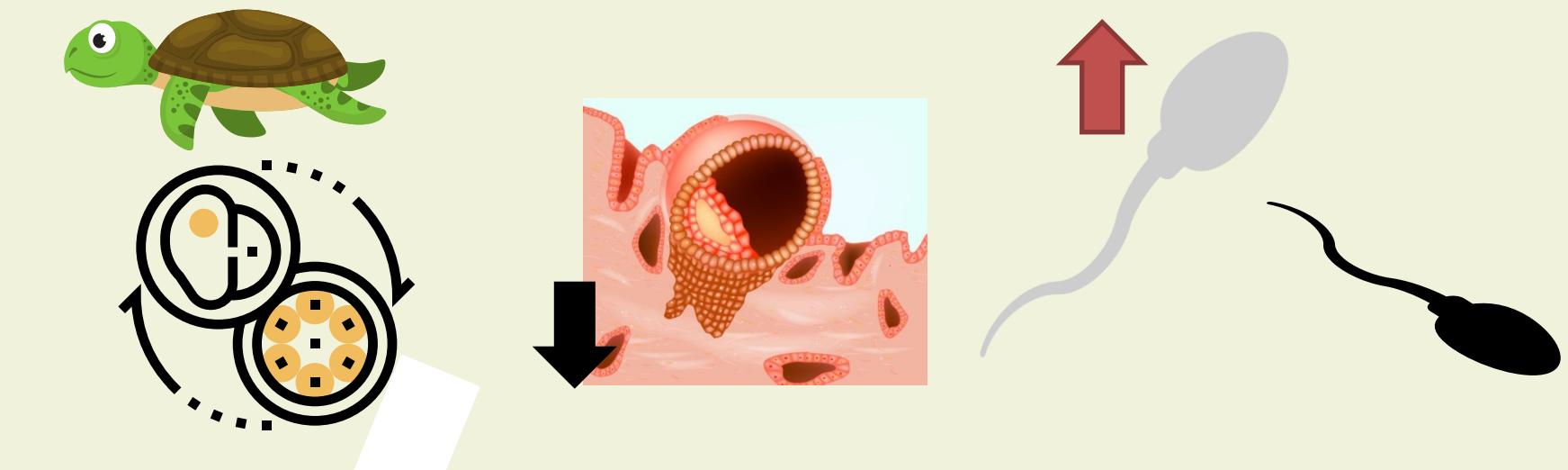


Observational Study ➤ Fertil Steril. 2019 Apr;111(4):699-707.e1.

doi: 10.1016/j.fertnstert.2018.11.035. Epub 2019 Feb 28.

Double-stranded sperm DNA damage is a cause of delay in embryo development and can impair implantation rates

Aida Casanovas ¹, Jordi Ribas-Maynou ², Sandra Lara-Cerrillo ², Ana Raquel Jimenez-Macedo ¹, Olga Hortal ¹, Jordi Benet ³, Joan Carrera ¹, Agustín García-Péiró ⁴

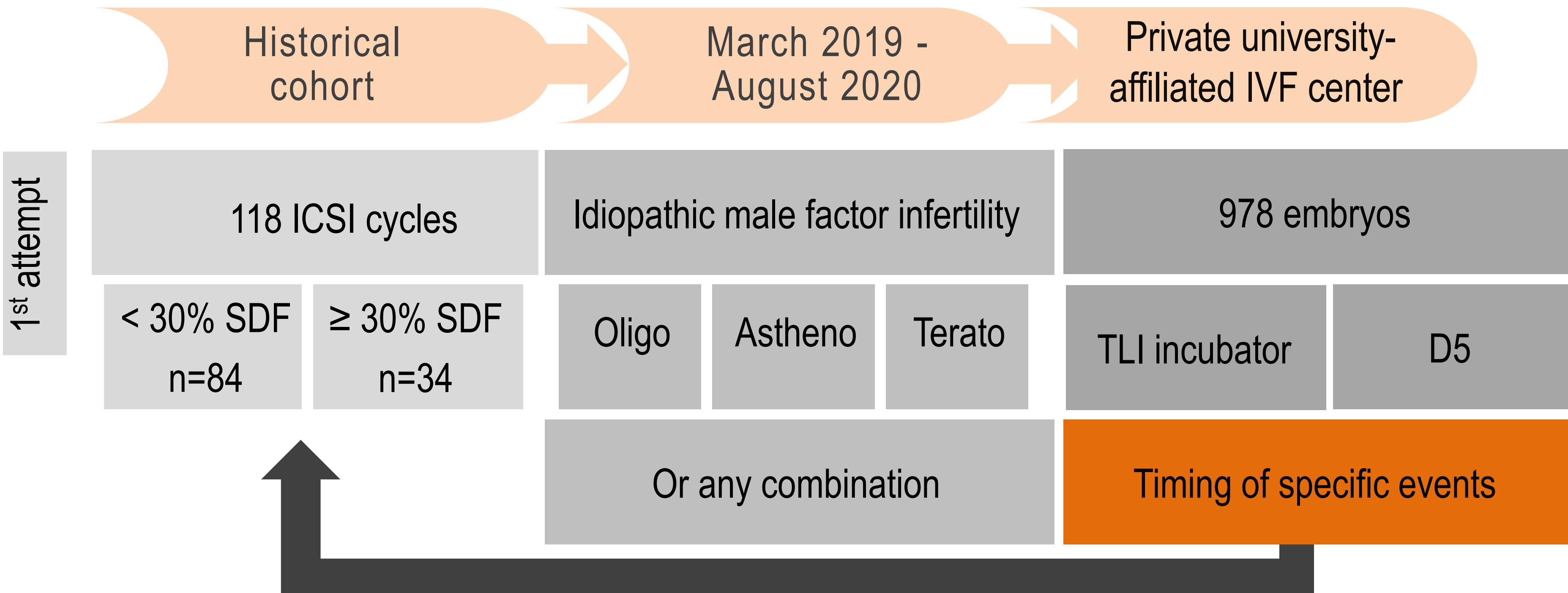


OBJECTIVE

To investigate if TLI can identify morphokinetic events impacted by high DFI, irrespective of conventional morphological embryo assessment and development rate

MATERIAL AND METHODS

Study design



MATERIAL AND METHODS

Controlled ovarian stimulation



GnRH Antagonist

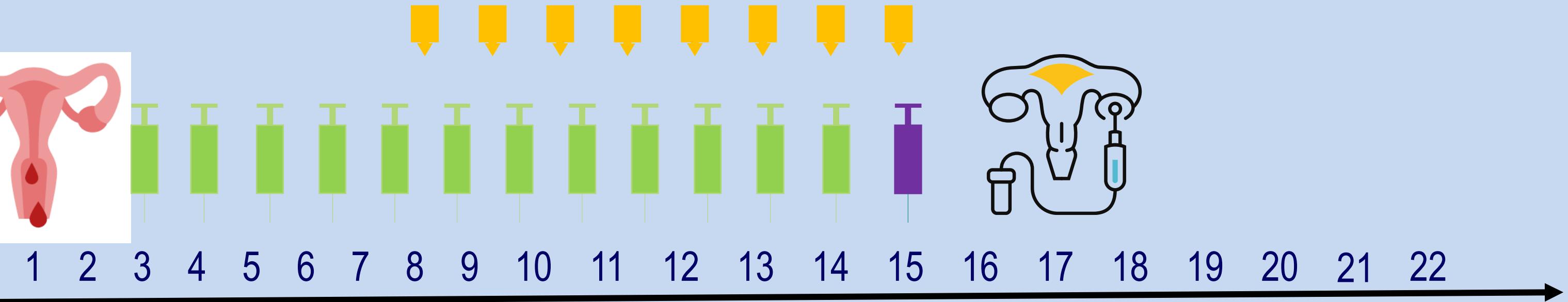
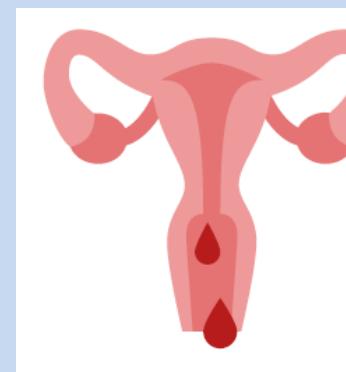
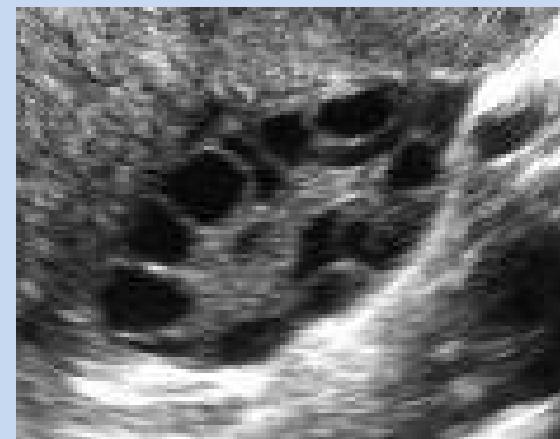


Recombinant FSH



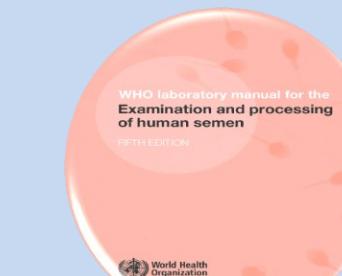
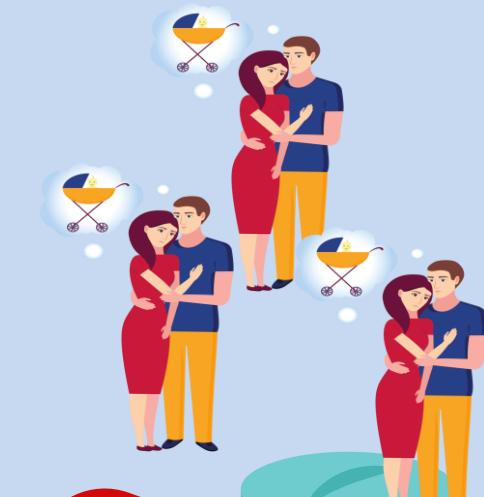
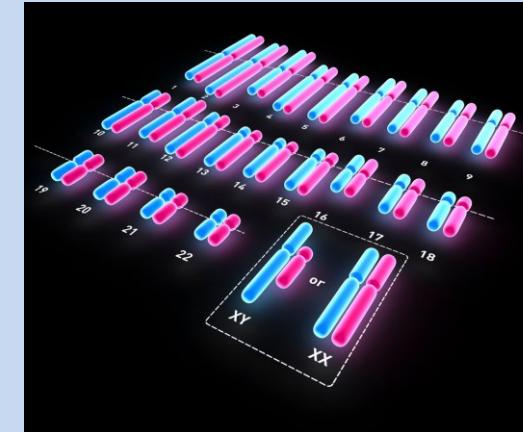
Recombinant hCG

E2

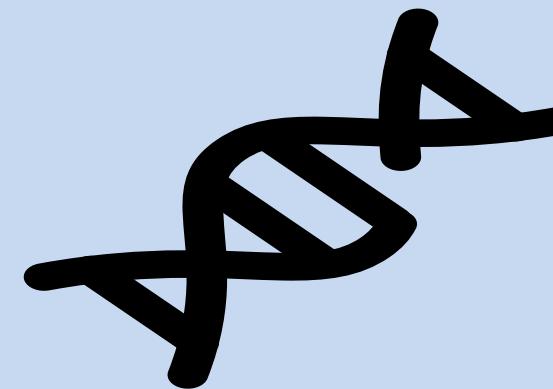


MATERIAL AND METHODS

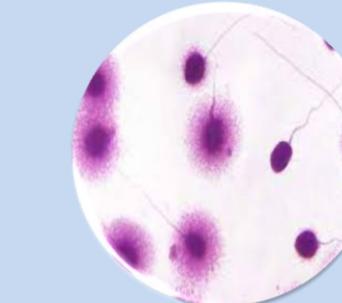
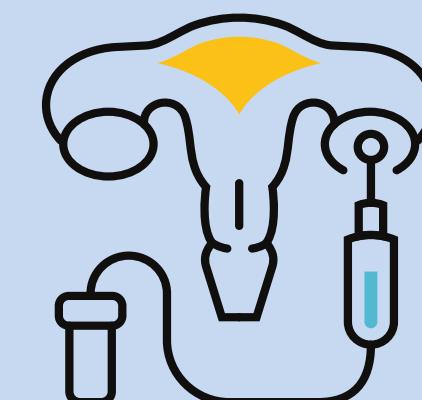
Male partner workup



WHO guidelines, 2010



2-layered density gradient centrifugation



Sperm chromatin dispersion (SCD) test

MATERIAL AND METHODS

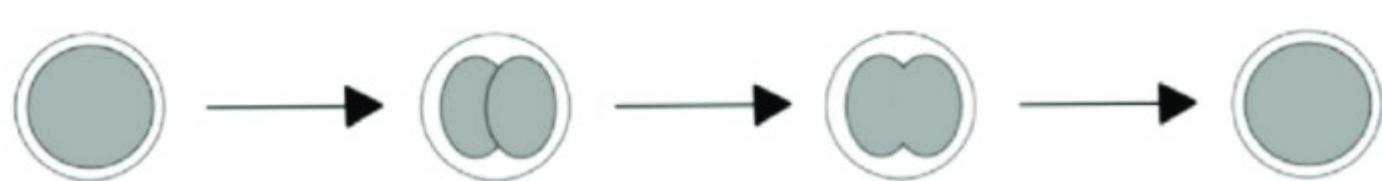
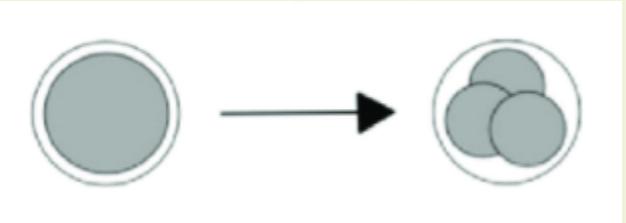
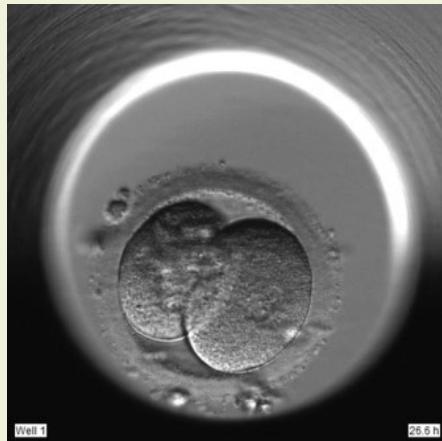
Embryo culture



Until day 5

11 focal planes

Every 10 minutes



Multinucleation

Abnormal cleavage patterns



One or two embryos transferred

MATERIAL AND METHODS

Data analysis and statistics

Post hoc power analysis



α 5%

GPower 3.1

527
blastocysts

Effect
 t_B

Superior to 80%



GzLM

Bonferroni post hoc

Timing of specific events

Mean \pm SD

OR with 95% CI

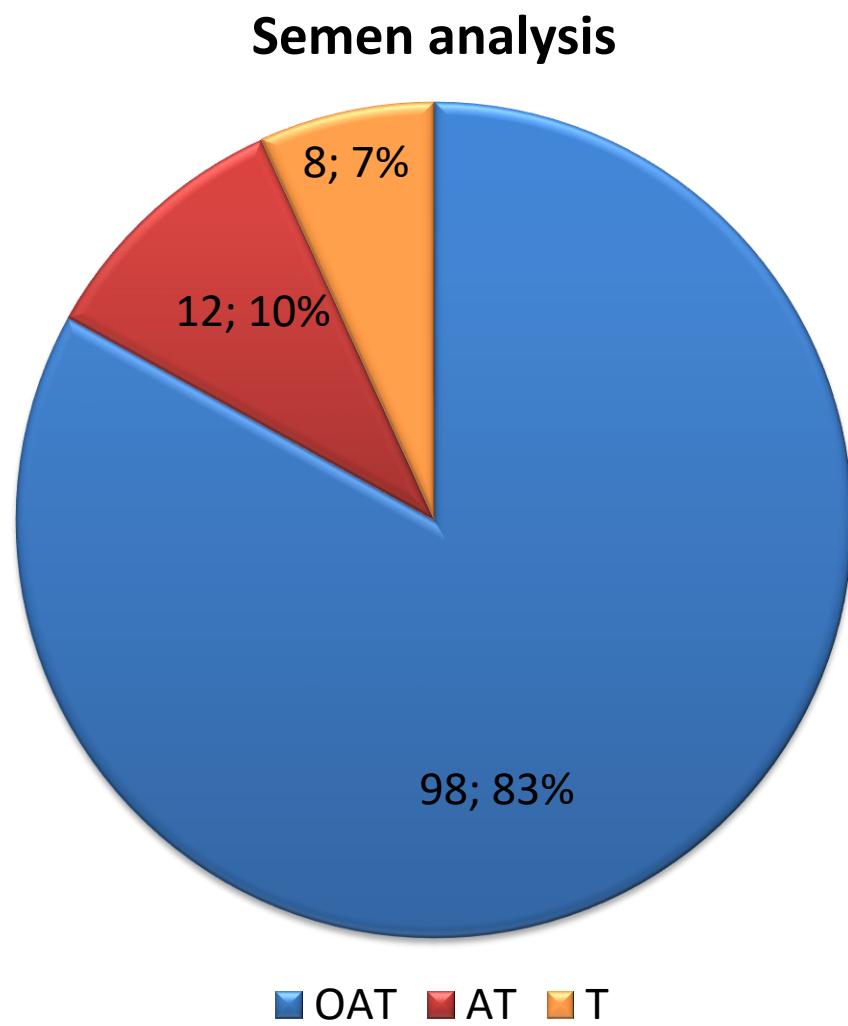
Independent: DFI groups

Dependent: kinetic markers

Covariates: maternal and paternal
ages, retrieved oocytes, EA

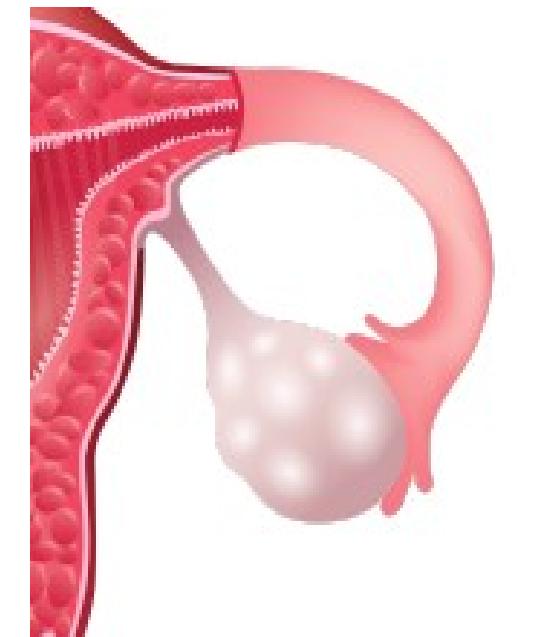
p-values (5%)

RESULTS



Variable	<30% DFI (n=84)	≥30% DFI (n=34)	p-value
Semen analysis			
Male age (years)	39.7 ± 5.0	38.8 ± 6.6	0.457
Ejaculatory abstinence length (days)	2.4 ± 0.9	2.6 ± 0.6	0.092
Sperm DFI (%)	18.2	42.8	<0.001
Semen volume (mL)			
Sperm concentration ($\times 10^6/\text{mL}$)	8.9 ± 12.7	9.1 ± 11.1	0.961
Sperm count ($\times 10^6$)			
	34.3 ± 87.9	24.7 ± 31.3	0.416

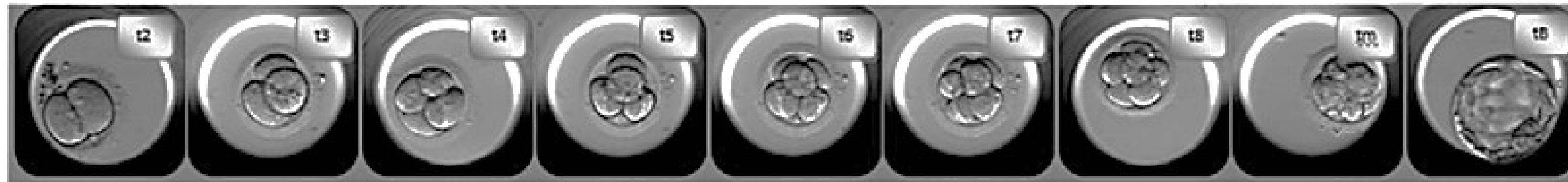
RESULTS



Variable	<30% DFI (n=84)	$\geq 30\%$ DFI (n=34)	p-value
Female age (years)	35.9 ± 4.2	36.9 ± 1.9	0.078
Female BMI	24.0 ± 3.3	24.6 ± 4.8	0.466
COS outcomes			
Total dose of FSH (IU)	2731.9 ± 905.7	2528.1 ± 726.9	0.265
IVF lab outcomes			
Estradiol level on hCG trigger (pg/mL)	2560.0 ± 1444.0	2442.1 ± 1983.1	0.804
Follicles (n)	15.8 ± 5.9	16.3 ± 9.6	0.754
Retrieved oocytes (n)	11.6 ± 5.7	11.3 ± 5.6	0.842
Mature oocyte rate (%)	78.5	82.4	0.195
Fortification rate (%)	73.0	76.6	0.416

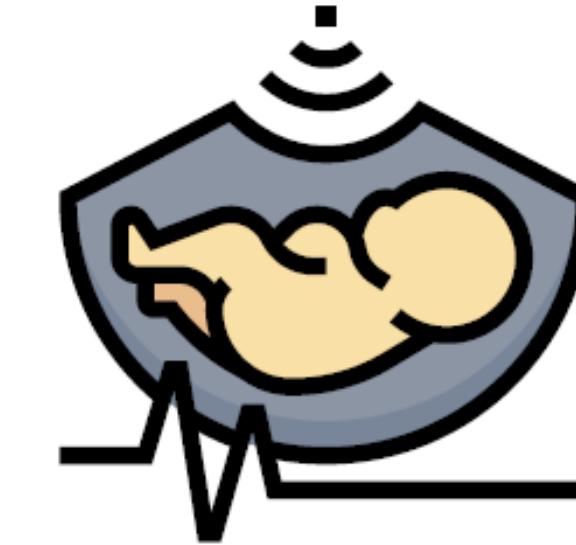
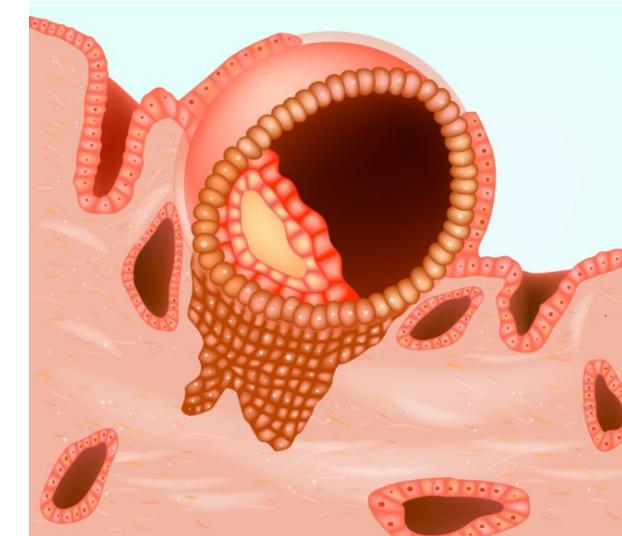


RESULTS



Morphokinetic data (hours)	<30% DFI (n=592)	≥30% DFI (n=386)	p-value
tPNa	6.1 ± 0.2	6.8 ± 0.2	0.030
tPNf	23.0 ± 0.3	24.2 ± 0.3	0.009
t2	25.4 ± 0.3	26.9 ± 0.3	0.002
t3	34.8 ± 0.3	37.3 ± 0.4	<0.001
t4	37.5 ± 0.4	39.3 ± 0.4	0.003
t5	46.2 ± 0.5	49.5 ± 0.6	<0.001
t6	49.7 ± 0.5	52.8 ± 0.6	0.001
t7	52.4 ± 0.6	55.6 ± 0.7	0.001
t8	56.2 ± 0.7	58.9 ± 0.8	0.017
tSB	97.5 ± 1.5	105.9 ± 1.7	0.002
tB	108.6 ± 0.8	112.4 ± 1.2	0.016

RESULTS

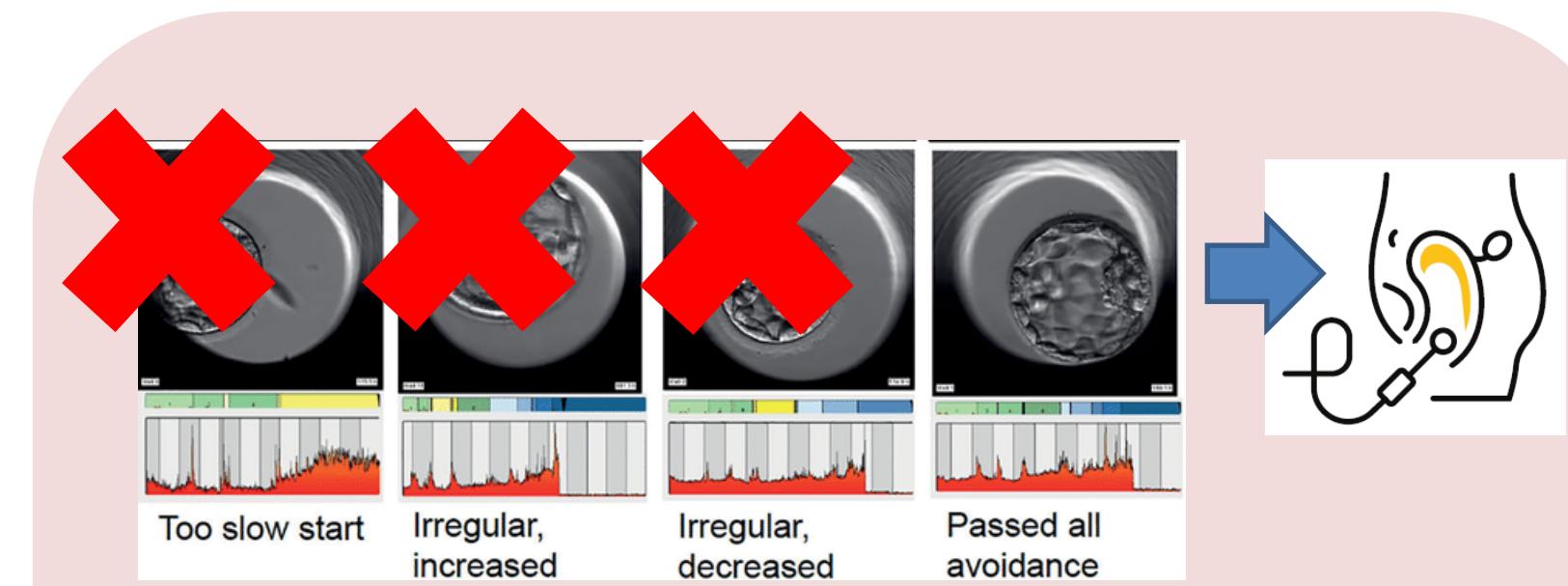


Variable	<30% DFI (n=84)	≥30% DFI (n=34)	p-value
<i>Clinical outcomes</i>			
Transferred embryos (n)	1.5 ± 0.5	1.5 ± 0.5	0.775
Implantation rate (%)	25.6	25.0	0.922
Pregnancy rate (%)	40/70 (57.1)	14/25 (56.0)	0.921
Miscarriage rate (%)	7/40 (17.5)	3/14 (21.4)	0.708

WIDER IMPLICATIONS OF THE FINDINGS

The findings presented here highlight the importance of TLI in the presence of high sperm DFI.

Variable	<30% DFI	≥30% DFI
Blastocyst development (%)	53.1	55.1
High-quality blastocyst rate (%)	87.9	86.2



Clinical outcomes (%)	<30% DFI	≥30% DFI
Implantation rate	25.6	25.0
Pregnancy rate	57.1	56.0
Miscarriage rate	17.5	21.4

› Hum Reprod. 2011 Oct;26(10):2658-71. doi: 10.1093/humrep/der256. Epub 2011 Aug 9.

The use of morphokinetics as a predictor of embryo implantation

Marcos Meseguer ¹, Javier Herrero, Alberto Tejera, Karen Marie Hilligsøe, Niels Birger Ramsing, Jose Remohí

CONCLUSION

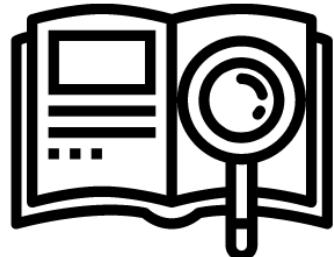
Embryo morphokinetic parameters are negatively impacted by high sperm DFI, resulting in delayed cell cleavage and blastulation, whereas conventional morphological embryo assessment and blastocyst development rate are not related to DNA integrity

STAFF



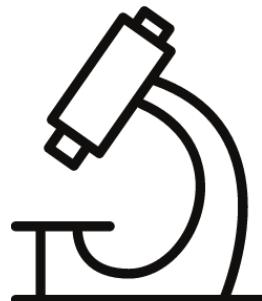
Clinical Board

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Edson Borges Junior



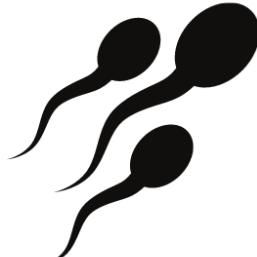
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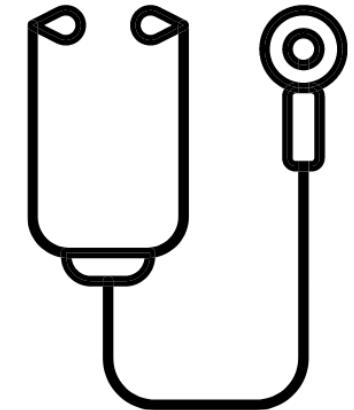
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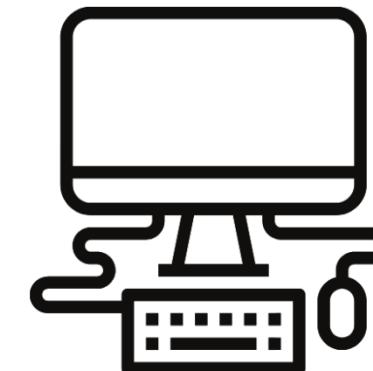
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Mauro Bibancos De Rose
Natalia Grandini Tannous
Paula Ferreiro Vieira



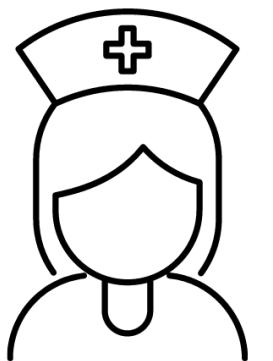
Support

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Simone de S. Carvalho
Leonardo S. Lopes



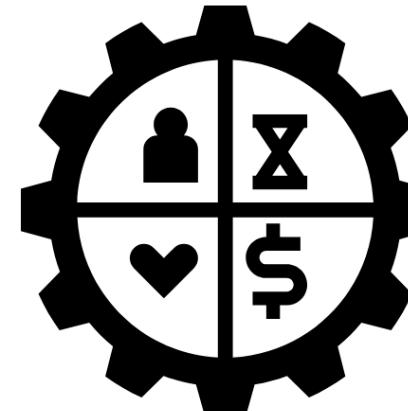
Information Technology

Marcelo Alexandre Baptista



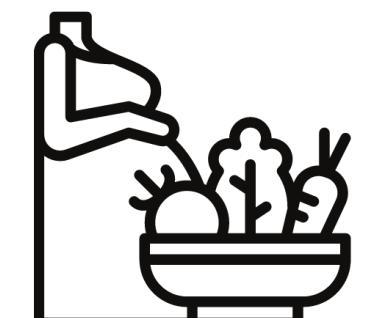
Nursing Team

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